

Network-based Analysis of Genome Wide Association Data Provides Novel Candidate Genes for Lipid and Lipoprotein Traits*[§]

Amitabh Sharma^{‡¶§§}, Natali Gulbahce^{¶¶**}, Samuel J. Pevzner^{¶¶|||}, Jörg Menche^{¶¶}, Claes Ladenvall[‡], Lasse Folkersen^{‡‡}, Per Eriksson^{‡‡}, Marju Orho-Melander^{‡§§}, and Albert-László Barabási^{¶¶|||}

Genome wide association studies (GWAS) identify susceptibility loci for complex traits, but do not identify particular genes of interest. Integration of functional and network information may help in overcoming this limitation and identifying new susceptibility loci. Using GWAS and comorbidity data, we present a network-based approach to predict candidate genes for lipid and lipoprotein traits. We apply a prediction pipeline incorporating interactome, co-expression, and comorbidity data to Global Lipids Genetics Consortium (GLGC) GWAS for four traits of interest, identifying phenotypically coherent modules. These modules provide insights regarding gene involvement in complex phenotypes with multiple susceptibility alleles and low effect sizes. To experimentally test our predictions, we selected four candidate genes and genotyped representative SNPs in the Malmö Diet and Cancer Cardiovascular Cohort. We found significant associations

with LDL-C and total-cholesterol levels for a synonymous SNP (rs234706) in the cystathionine beta-synthase (CBS) gene ($p = 1 \times 10^{-5}$ and adjusted- $p = 0.013$, respectively). Further, liver samples taken from 206 patients revealed that patients with the minor allele of rs234706 had significant dysregulation of CBS ($p = 0.04$). Despite the known biological role of CBS in lipid metabolism, SNPs within the locus have not yet been identified in GWAS of lipoprotein traits. Thus, the GWAS-based Comorbidity Module (GCM) approach identifies candidate genes missed by GWAS studies, serving as a broadly applicable tool for the investigation of other complex disease phenotypes. *Molecular & Cellular Proteomics* 12: 10.1074/mcp.M112.024851, 3398–3408, 2013.

Genome wide association studies (GWAS)¹ meta-analyses have pinpointed a number of new gene regions contributing to multifactorial diseases. GWAS typically find limited numbers of loci that contribute modestly to complex phenotypes (1), and GLGC meta-analysis of GWAS data has reached the limit of what can be expected (2) without the use of alternative strategies. Given that susceptibility loci for complex traits are unlikely to be randomly distributed in the genome (3), we might expect that the genes associated with a disease will be more likely to be present within the same pathways or functional groupings. In published cases, pathway based GWAS analysis provides an alternative approach to the dissection of complex disease traits (4, 5). In addition, nominal GWAS p values superimposed upon the human molecular network have been used to identify genes associated with multiple sclerosis (6), and the disease association protein–protein link evaluator (DAPPLE) has been used to find significant interactions among proteins encoded by genes in loci associated

From the [‡]Department of Clinical Sciences, Diabetes and Cardiovascular Disease, Genetic Epidemiology, Lund University, University Hospital Malmö, Malmö, Sweden; [§]Center for Complex Network Research and Department of Physics, Northeastern University, Boston, Massachusetts 02115, USA; ^{¶¶}Center for Cancer System Biology (CCSB) and Department of Cancer Biology, Dana-Farber Cancer Institute and Department of Genetics, Harvard Medical School, 44 Binney Street, Boston, Massachusetts; ^{|||}Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, 75 Francis Street, Boston, Massachusetts 02115; ^{**}Department of Cellular and Molecular Pharmacology, University of California 1700 4th Street, Byers Hall 308D, San Francisco, California 94158; ^{‡‡}Atherosclerosis Research Unit, Department of Medicine, Karolinska Institute, Stockholm, Sweden; ^{¶¶}Department of Biomedical Engineering, Boston University, Boston, MA 02215; ^{|||}Boston University School of Medicine, Boston, MA 02118

Received October 10, 2012, and in revised form, July 22, 2013

Published, MCP Papers in Press, July 23, 2013, DOI 10.1074/mcp.M112.024851

Author Contributions: A.S. and N.G. designed, interpreted and wrote the manuscript. C.L. helped in analysis of genome wide association data for SNP annotation. J.M. and S.P. assisted in computing comparison analysis with different methods using GO terms, GWAS and functional data. L.F. and P.E. contributed data and analysis for CBS eQTL analysis. M.O.-M. has contributed in planning the project and helped in writing the final version of the manuscript. A.L.B. contributed in critical evaluation of the manuscript. All authors discussed the results and commented on the manuscript.

¹ The abbreviations used are: GWAS, Genome wide association studies; SNP, single nucleotide polymorphism; GO, Gene Ontology; GCM, GWAS-based-meta analysis Comorbid Module; GLGC, Global Lipids Genetics Consortium; MT, molecular triangulation; KEGG, Kyoto Encyclopaedia of Genes and Genome; BIGG, biochemically, genetically, and genomically structured genome scale metabolic network reconstruction; eQTL, Expression quantitative trait loci.

with other particular diseases (7). Other approaches incorporate heterogeneous molecular data such as linkage studies, cross species conservation measures, gene expression data and protein–protein interactions to better understand GWAS results (8, 9). Integrating molecular network information, pathway analyses, and GWAS data thus holds promise for identifying new susceptibility loci and improving the identification of relevant candidate genes.

If a gene is involved in a specific functional process or disease, its molecular network neighbors might also be suspected to have some role (3). In line with this “local” hypothesis, proteins involved in the same disease show a high propensity to interact (10) or cluster together (11) with each other. Interactions between variations in multiple genes, each with strong or modest effects, perturbing the same pathways or modules, may govern complex traits (3, 6). The molecular triangulation (MT) algorithm can be applied to rank seed genes according to their common disease associated neighbors, assigning closer and more connected neighbors higher values (12). Interactions between modestly associated MT genes may be indicative of coherent disease pathways or of genes conferring susceptibility to disease in a coordinated manner. The *jActiveModule* method (13) combines seed gene scores with biologically relevant interactions to identify network modules where perturbations causative of disease are more likely to reside. Lastly, although not yet implemented at the module level, phenotypic coherence between interacting pairs of genes has been quantified using the combination of molecular level gene to disease relationships and Medicare comorbidity data (14, 15).

We believe that GWAS significant SNPs and variants representing potential candidate genes can use the above strategies to reveal more about the missing heritability of complex phenotypes. The most important risk factors for coronary artery disease (CAD) include serum concentrations of total cholesterol (TC), low-density lipoprotein cholesterol (LDL-C), high-density lipoprotein cholesterol (HDL-C) and triglycerides (TG). We present a GWAS-based meta-analysis Comorbid Module (GCM) approach that uses significant ($p < 5 \times 10^{-8}$) GWAS signals for these four traits in the context of molecular networks to prioritize modules of disease-associated candidate genes. We evaluate our approach experimentally through allelic association and genotyping within the Malmö Diet and Cancer Cardiovascular Cohort (MDC-CC) for SNPs representing top candidate genes.

MATERIALS AND METHODS

The GWAS Comorbid Module (GCM) approach to predict lipid/lipoprotein trait candidate genes involves the following steps:

- (1) Mapping of GLGC GWAS meta-analysis SNPs to genes.
- (2) Construction of a human interactome, pooling protein interaction data from different sources.
- (3) Identification of candidate genes associated with lipid/lipoprotein traits using molecular triangulation (MT).

(4) Identification of modules of seed and neighboring genes using the *jActiveModule* method (jAM).

(5) Selection of phenotypically coherent (GCM) modules of seed and candidate genes using comorbidity analyses.

(6) Validation of pipeline outputs (MT, jAM, and GCM) and comparison to other methods (CANDID and MetaRanker).

(7) Selection of SNPs representing GCM candidate genes for genotyping in the MDC-CC.

In summary, we curate GWAS-based seed genes ($p < 5 \times 10^{-8}$), constructed an interactome, implement the MT method, filter MT candidate genes by *jActiveModule* results, select phenotypically coherent modules, validate the outputs of every step, and genotype SNPs representing GCM candidate genes of interest for lipid and lipoprotein traits.

(1) *Mapping of GLGC GWAS Meta-analysis SNPs to Genes*—The GLGC GWAS meta-analyses data is based on 46 lipid/lipoprotein GWAS involving over 100,000 individuals of European descent as ascertained in the United States, Europe, or Australia (16). The GLGC consortium contributed genome wide analysis data for analyses, including ~2.6 million genotyped or imputed SNPs associated with four traits (TC, LDL-C, HDL-C and TG). The entire set of HapMap phase III SNPs and pairwise LD estimates (Release 27) for the CEU population was downloaded, and LD pruning and SNP to gene mapping was performed as described previously (3). If a SNP could be mapped to more than one gene, all genes were included, and SNPs located in gene desert regions were excluded from our analysis. To be sure of the robustness of our results, we also annotated SNPs using the ProxyGeneLD tool (17) and found similar SNP to gene annotations. Genes representing SNPs with GWAS significant p values of less than 5×10^{-8} were used as “seed” genes.

(2) *Construction of a Human Interactome, Pooling Interaction Data From Different Sources*—We created a human interactome consisting of proteomic, transcriptional, and metabolic interactions. Protein–protein interactions from three high-throughput yeast-two-hybrid datasets were combined with the binary subset of interactions reported in the IntAct and MINT databases (18–22). Together, these data sets describe 15,315 interactions between 6101 gene-coding proteins. For regulatory interactions, we used the TRANSFAC database version 2008.2, which included 1340 links between 271 human transcription factors and their 564 targets (23). Metabolic coupling interactions were derived from the Kyoto Encyclopedia of Genes and Genomes (KEGG) and the Biochemically, Genetically, and Genomically structured genome scale metabolic network reconstruction (BiGG) database as described in (15); 10,642 such metabolic links between 921 enzymes were included. The union of these sets of interactions yielded an interactome of 7117 (N) proteins and 21,964 (M) links, with an average shortest path length $\langle l \rangle$ of 4.52.

(3) *Identification of Candidate Genes Associated With Lipid/Lipoprotein Traits Using Molecular Triangulation (MT)*—MT begins with sets of seed (disease) genes known to be associated with phenotypes and suggests additional disease genes, typically network neighbors of multiple seed genes (12, 24). Here, we used the strengths of GWAS signals for lipoprotein traits and SNP to gene mappings to assign primary evidence scores to seed genes. MT used these primary evidence scores and the position of these seed genes within the interactome to calculate secondary evidence scores for neighbors (12). To calculate the significance of the secondary evidence scores, we performed 1000 degree-preserving network randomizations. The significance of the scores was then calculated as described by Iosifov *et al.* (supplementary material) (24). We applied Benjamini and Hochberg false discovery rate (FDR) corrections with a 0.05 FDR threshold to our predictions; this meant that we expected less than 5% of our predictions to be false positives.

(4) *Identification of Modules of Seed and Neighboring Genes Using the jActiveModule Method (jAM)*—The MT method results in a large number of statistically significant predictions, but some of the predictions may be artifacts of low or excessive connectivity (24). To address this concern, we independently implemented the *jActiveModule* method to determine modules with maximal proportions of the lowest *p* value genes. Later we pruned the MT gene sets to only include genes that were within these modules.

The *jActiveModule* method uses GWAS association *p* values of seed genes and interactome context to produce aggregated module scores. The method compares real network modules to those derived from 10,000 matched randomized network Monte-Carlo simulations (13). To examine the effect of the GWAS signal strength distribution by itself, we compared the real module scores to distributions based on randomized gene to trait association *p* values. Matched numbers of seed genes were chosen from the set annotated by NCBI (Ver. 36) and from the set described by the Online Mendelian Inheritance in Man database (OMIM, December 2009 release). The differences between outputs after either randomization strategy are described in supplementary material.

As additional controls for the *jActiveModule* results, we implemented the Molecular COmplex DETection (MCODe) algorithm, the Markov Clustering algorithm (MCL), and the Klein-Ravi Steiner tree algorithm with submodule detection using MCL (GenRev package (25)). Parameters for the MCL and MCODe algorithms were adopted from a previous study (26). To compare the results from different approaches, we used the Jaccard similarity (*J*) between the sets of seed genes (*S*) and putative module genes (*T*) determined using each method:

$$J = \frac{|S \cap T|}{|S \cup T|} \quad (\text{Eq. I})$$

where, $|S \cap T|$ is the intersection of sets *S* and *T* and $|S \cup T|$ is the union.

(5) *Selection of Phenotypically Coherent (GCM) Modules of Seed and Candidate Genes Using Comorbidity Analyses*—To further rank the modules, we used OMIM gene-disease associations to perform analyses of comorbidity based on the co-occurrence of ICD-9 codes taken from a 13 million patient Medicare data set (14). OMIM diseases were manually mapped to ICD9 codes so that interactions between genes could be supported by comorbidity between their associated diseases. To quantify comorbidity, Relative Risk (RR) scores were calculated for every pairwise combination of diseases associated with at least one of the genes in the module:

$$RR = \frac{C_{12} \times np}{P_1 \times P_2} \quad (\text{Eq. I})$$

C_{12} = number of patients who had both disease 1 and 2

P_1 = number of patients who had disease 1

P_2 = number of patients who had disease 2

np = 13,039,018 (total number of patients)

Lower and upper bounds (*lb* and *ub*) of 99% confidence intervals were calculated according to the Katz *et al.* method (24):

$$lb = RR \times \exp(-2.576 \times \sigma) \quad (\text{Eq. II})$$

$$ub = RR \times \exp(2.576 \times \sigma) \quad (\text{Eq. III})$$

where σ is given by:

$$\sigma = \sqrt{\frac{1}{C_{12}} + \frac{1}{P_1 \times P_2} - \frac{1}{np} - \frac{1}{np \times np}} \quad (\text{Eq. IV})$$

The relative risk was taken to be significant when the 99% confidence interval did not include the expected value of one, which would indicate findings of no consequence. To summarize the pairwise

comorbidities for each module and rank them, we averaged the pairwise RR scores between associated ICD9 disease codes, creating a module relative risk (mRR) score. A *Mann-Whitney U* test was used to compare the observed mRR scores to those of 100 randomly constructed modules.

(6) *Validation of Pipeline Outputs (MT, jActiveModule and GCM) and Comparison to Other Methods (CANDID and MetaRanker)*—We validated the MT, *jActiveModule* and GCM steps using data from Teslovich *et al.* (1) as a benchmarking set. Several measures of predictive power were used: (1) precision $[TP/(TP+FP)]$, (2) specificity $[TN/(TN+FP)]$, and (3) accuracy $[(TP+TN)/(TP+FP+FN+TN)]$, where TP is number of true positives or candidate genes correctly identified as disease genes, TN is number of true negatives or correctly identified nondisease genes, FP is number of false positives or nondisease genes identified as candidate genes, and FN is number of false negatives of disease associated genes that were not identified as candidates. We evaluated the functional coherence of candidate genes relative to seed genes by comparing their enrichment, as a set, for functional annotations. These allowed us to evaluate the consistency of candidate gene sets with respect to phenotypically similar diseases (27). After determining the GO biological process terms enriched within the sets of seed genes, we tested the enrichment of these terms in candidate genes.

To perform a comprehensive comparison of GCM to other methods, we implemented MetaRanker (8) and CANDID (9). MetaRanker predicts candidate genes by integrating complementary layers of protein interaction, linkage, GWAS, differential expression and disease data. These five different data sources are integrated into a single meta-evidence rank, quantifying the likelihood of genes being involved in a disease of interest (8). CANDID is designed to rank candidate genes by eight evaluation criteria, considering associated publications, protein domains, conservation, expression, interactions, linkages, SNP associations, and custom data (9). We compared the top 200 MetaRanker and CANDID candidate genes to the outputs of the MT, *jActiveModule* and comorbidity analysis steps. We then benchmarked candidate gene sets sharing GO terms with the seed genes, computing precision, specificity and accuracy as described previously for the candidate genes from each step of the GCM method as well as the CANDID and MetaRanker outputs.

In addition, using GeneWanderer, we tested how parsimoniously candidate genes from the GCM approach were involved in obesity, which is known to be related to lipid/lipoprotein traits. GeneWanderer was provided with genomic locations 1Mb in either direction of SNPs representing GCM genes and ranked candidate genes within these windows according to their single shortest path through the STRING (28) network to obesity genes.

(8) *Selection of SNPs representing GCM candidate genes for genotyping in the MDC-C*—GCM genes with the strongest co-expression correlations to the seed genes were selected for genotyping. Genome wide mRNA expression data of 79 human tissues were obtained from the Gene Expression Atlas (29). Spearman's test was used to assess correlation between GCM gene and seed gene mRNA expression, with the criterion for significance set at $\rho > 0.5$ and $p < 0.05$. The second criteria considered was sequence conservation of the regions in which SNPs were located, as alterations at conserved sites have more drastic functional effects when changed (30). The third criteria considered was the position of the SNPs relative to the candidate genes. Among conserved SNPs with GLGC GWAS *p* values < 0.05 , we used following hierarchy to rank importance for further genotyping: coding > intronic > 5'UTR > 3'UTR > 5'upstream > 3'upstream (31).

Study Population: The Malmö Diet and Cancer Cardiovascular cohort (MDC-CC)—The Malmö Diet and Cancer (MDC) study is a community-based prospective cohort of 28,449 persons originally recruited for baseline examination between 1991 and 1996 (32, 33).

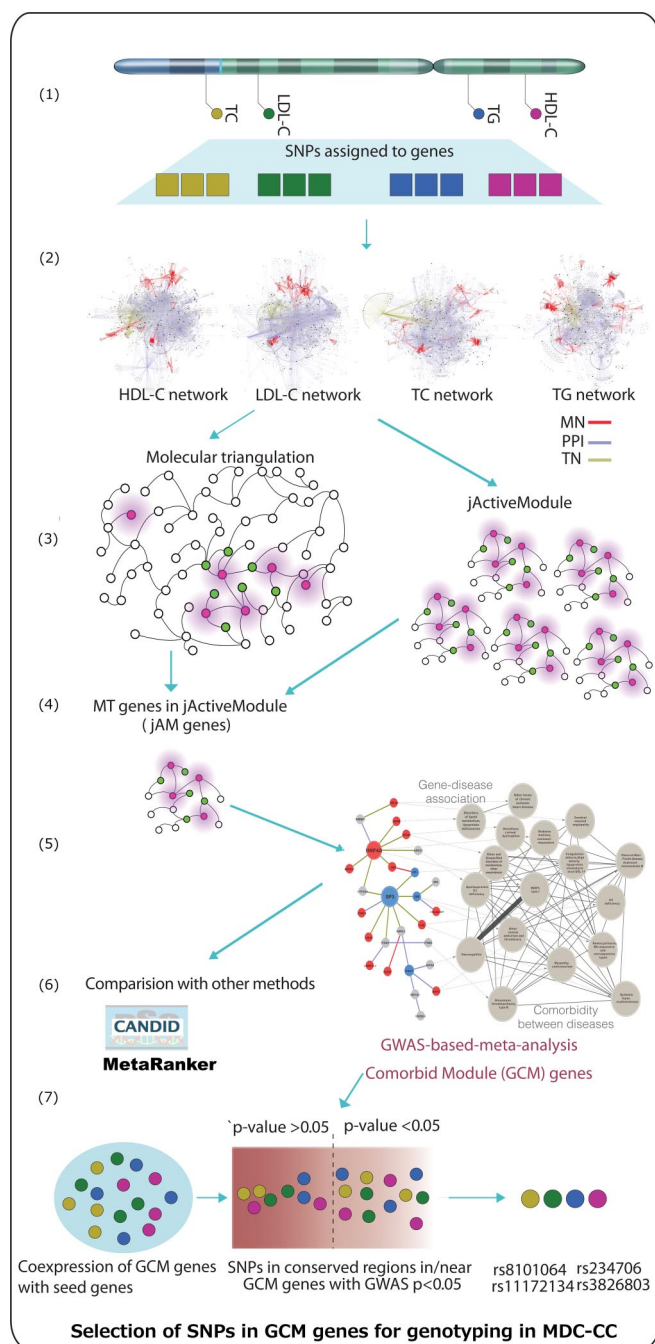


FIG. 1. Schematic representation of GCM approach. (1): Mapping of GLGC GWAS meta-analysis SNPs to genes. (2): Construction of a human interactome by pooling protein interaction data from different sources and mapping seed genes within the network. (3): Identification of candidate genes associated with lipid/lipoprotein traits using molecular triangulation (MT). (4): Identification of seed and neighbouring gene modules using the jActiveModule (jAM) method, pruning of MT candidate gene sets. (5): Selection of phenotypically coherent (GCM) modules of seed and candidate genes using comorbidity analyses. (6): Validation of MT, jAM and GCM gene set outputs and comparison to CANDID and MetaRanker methods. (7): Selection of SNPs, representing GCM candidate genes, for genotyping in the MDC-CC. GCM genes were prioritized based on their co-expression

From this cohort, 6103 persons were randomly selected to participate in the Cardiovascular Cohort (MDC-CC), which seeks to investigate risk factors for cardiovascular disease. All participants underwent questioning regarding their medical history, a physical examination, and a laboratory assessment for cardiovascular risk factors. In fasting venous blood samples, TC, HDL-C, and triglyceride levels were measured according to standard procedures by the Department of Clinical Chemistry at University Hospital Malmö. Levels of LDL-C were calculated according to Friedewald's formula, with the assignment of missing values to subjects with a triglyceride level of more than 4.5 mmol per liter. DNA was available from 5763 individuals for genotyping, and of these individuals, lipid levels were available for 5056 individuals that were not on lipid lowering medication. The ethics committee of Lund University approved the MDC-CC study protocols, and all participants provided written informed consent.

Genotyping—Genotyping of the selected SNPs was performed using genomic DNA from 5763 individuals using the allelic discrimination method using an ABI 7900 instrument (Applied Biosystems, Foster City, CA). Samples that were successfully genotyped for at least 50% of the SNPs were included in further analyses ($n = 5698$). We confirmed that the genotypes were at Hardy-Weinberg equilibrium. The overall genotyping success rate was 98.2%. For this epistasis analysis, we created a variable indicating how many risk alleles (increasing total cholesterol, LDL-cholesterol or triglycerides and/or lowering HDL cholesterol) each individual in the population cohort was carrying, *i.e.* summing up the number of risk-alleles to a variable “risk-allele score.” For the four SNPs, the theoretical maximal number of risk-alleles was eight (for individuals homozygous for risk alleles of all four SNPs) and the minimum was zero. In MDC-CC cohort, individuals ranged from zero to six risk alleles, and the risk-allele score was used as a variable in a linear regression analysis adjusting for age and sex to analyze if the combined effect of the four SNPs resulted in an association with lipid levels.

Expression Quantitative Trait (eQTL) Analysis—RNA extracted from the livers of 206 patients undergoing aortic valve surgery and/or surgery for aortic aneurysms (34) was hybridized to Affymetrix ST 1.0 Exon arrays. DNA extracted from circulating blood cells was hybridized to Illumina 610w-Quad BeadArrays. The association was tested using a linear additive model with corrections for age and gender. The average age of patients was 63.9 ± 11.8 years, with average total cholesterol levels of 5.05 ± 1.09 mm and average LDL-C levels of 3.11 ± 0.93 mm. None of the patients were known to have liver disease.

Gene Set Enrichment for Biological Pathways—To find statistically over-represented Gene Ontology (GO) annotations for candidate genes at each of the analysis steps, we used the Biological Networks Gene Ontology tool (BiNGO) implemented in Cytoscape. Enrichment analyses were performed using a hyper-geometric test followed by a Benjamini and Hochberg multiple hypothesis correction with a 0.05 FDR threshold (35). Odds ratios to measure the magnitudes of the enrichment were calculated using raw BiNGO outputs.

RESULTS

Introducing a network-based integrative approach, we identified novel candidate genes for lipid and lipoprotein traits (Fig. 1).

Prediction of Lipid/Lipoprotein Trait Candidate Genes Using Molecular Triangulation (MT)—We used the MT method to

with seed genes and hierarchical criteria including the genomic locations of SNPs, if the SNPs were synonymous variations, if the SNPs were in conserved regions of the genome, and GLGC GWAS-meta-analysis p -values ($p < 0.05$).

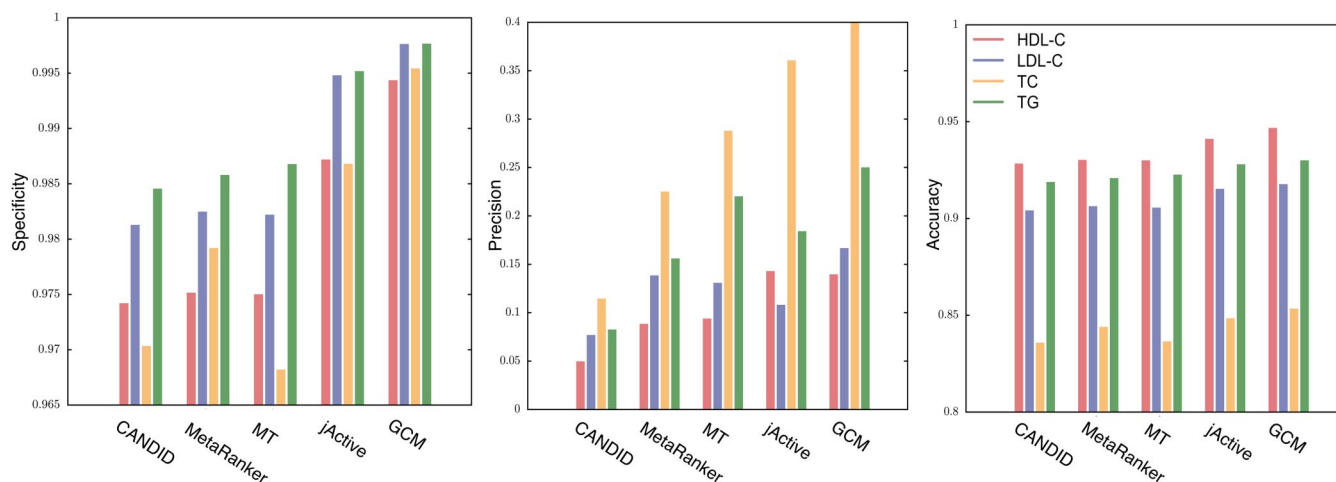


FIG. 2. Performance of CANDID, MetaRanker, MT, *jActiveModule* and GCM with respect to benchmarking dataset. The histograms show an increase in specificity, precision and accuracy with each of the steps.

identify phenotypically related candidate genes (MT-genes) based on their proximity within the interactome to seed genes associated with HDL-C, LDL-C, TG and TC traits (supplemental Table S1). The MT method had an accuracy of 98% in classifying true positives and true negatives for the four traits, with 33% precision and 98% specificity (Fig. 2).

Refinement of Candidate Gene Sets Using the *jActiveModules Method*—Although direct interactions can be used to identify candidate genes, modules in biological networks represent connected components contributing to cellular functions in a coordinated manner. Disruptions of these modules, which include both identified and unidentified disease genes, result in disease phenotypes (36). To avoid method specific biases, we implemented the *jActiveModule* algorithm in parallel with MT and used the intersection of the two result sets to identify more cohesive modules of seed and MT genes. Comparison of MT and *jAM* genes, based on their degree of connectivity, indicated that only retaining genes within the intersection of the two groups removed a significant number of the nodes having only one or greater than one hundred connections. ($p = 0.0017$, odds ratio = 2.01, supplementry Fig. S1). Furthermore, after the *jActiveModule* filtering step, precision increased from 33% to 44%, with 99% specificity and accuracy (Fig. 2).

The *jActiveModule* method uses GWAS association p values of seed genes within the interactome context to produce aggregated module scores, and these scores determined the extent to which randomized inputs could create coherent modules. Confirming the usefulness of the network context using randomized controls, the top 20 *jActiveModule* subnetwork scores were significantly higher than those from 100 randomization controls for each of the four traits ($p \leq 0.001$, supplemental Fig. S2).

Compared with the Steiner tree-MCL approach (supplemental Fig. S3), the *jActiveModule* algorithm identified modules with greater percentages of seed genes. Gene sets iden-

TABLE I
Mean relative risk (mRR) scores of the network modules. TC trait had the highest mRR score among the four. mRR-score is given as mean \pm S.D.

Trait	Number of modules	mRR-Score	* p value
HDL-C	157	1.8 \pm 1.0	~ 0.002
LDL-C	64	2.9 \pm 2.5	~ 0.001
TC	89	2.8 \pm 2.4	0.03
TG	85	1.8 \pm 0.6	~ 0.001

tified by *jActivemodule* were smaller and localized more tightly around seed genes within the interactome. This was likely because of *jActivemodule*'s flexible search for multiple modules, as opposed to the Steiner tree based method's strategy of attempting to find a single module connecting all of the seed genes in the interactome (37). Comparison with conventional clustering methods such as MCL and MCODE suggest that these methods are more suited to the identification of individual protein complexes (37) as well, while the *jActiveModule* method is more suited to the identification of multiple modules of seed genes spread throughout the interactome.

Retention of Modules According to Comorbidity Analyses Using Medicare Data (GCM)—To determine those modules with the most phenotypically coherent associations, we quantified the strengths of comorbidities between diseases associated with their genes. We implemented mRR scores, as explained in the methods section, because we believed that co-occurring diseases might be driven by related molecular machinery. We found 48 comorbid modules with mRR scores higher than one for HDL-C, (mean mRR score of 1.8, average $P \sim 0.002$), 15 modules for LDL-C, (mean mRR score of 2.9, average $P \sim 0.001$), 15 modules for TC, (mean mRR score of 2.8, average $P \sim 0.05$ and 23 modules for TG, (mean mRR score of 1.8, average $P \sim 0.001$). (Table I). Filtering the modules to only include those with above average mRR scores, precision increased from 44% to 55%, with 99% specificity

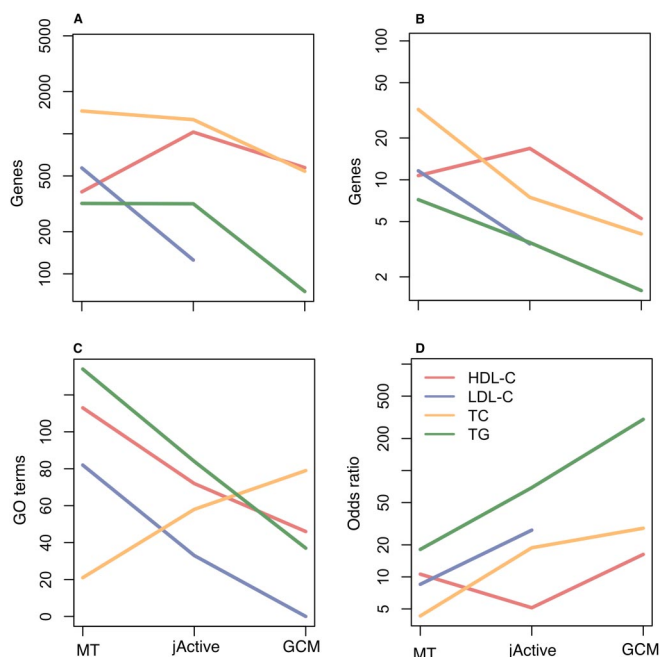


FIG. 3. GO term enrichments for gene sets. A, Mean numbers of genes, anywhere in the genome, associated with GO terms for which significant enrichments were found. B, Mean numbers of genes, within the sets of genes being tested, found to be associated with GO terms for which the gene sets were enriched. C, Counts of GO terms for which sets of candidate genes were enriched. D, Median odds ratios of GO term enrichment as a measure of enrichment effect size.

and accuracy (Fig. 2). ICD-9 codes associated with the retained GCM modules in all four of the primary traits included lipid metabolism, carbohydrate and transport metabolism, amino acid transport metabolism, being overweight, essential hypertension, cardiomyopathy, symptoms concerning nutrition, chronic ischemic heart disease, acute myocardial infarction, and diabetes mellitus (supplemental Fig. S4).

In addition to highlighting the phenotypic cohesiveness of the final GCM gene sets using ICD9 codes, the progressive benefit of the filtering steps was also quantified using GO term enrichment tests of the gene sets found at each step. We found that filtering MT genes by *jActive* module membership and only retaining modules with the most significant comorbidities yielded enrichments of progressively more specific GO terms annotating fewer genes (Fig. 3A). The cost of this was that the numbers of genes contributing to particular GO term enrichments decreased as genes were filtered away (Fig. 3B), and that except for the TG set of genes, fewer GO terms were detected (Fig. 3C). This filtered subset of more specific GO terms, however, displayed a trend of increasingly drastic effect size as measured by odds ratios of enrichment (Fig. 3D).

Validation of Pipeline Outputs (MT, jActiveModule and GCM) and Comparison to Other Methods (CANDID and MetaRanker)—Enrichment for GO term effect sizes (supplemental Fig. S5) and functional coherence of candidate and seed gene sets (Fig. 2) for the MT, *jActiveModule*, and GCM

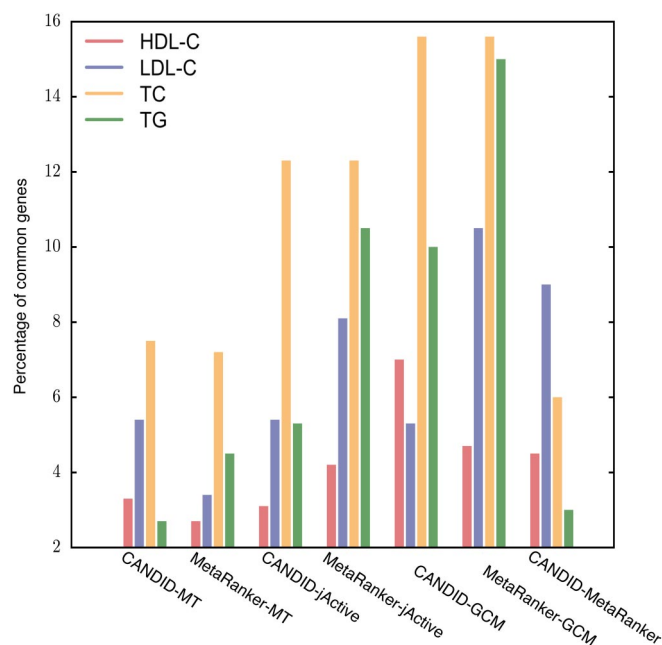


FIG. 4. Percentage of overlapping candidate genes between CANDID or MetaRanker and each of the GCM steps.

methods was greater than that of MetaRanker (8) and CANDID (9). Comparing overlaps between the candidate gene sets (Fig. 4), we found that predictions from CANDID and MetaRanker had 9% ($p < 0.0001$) overlap with each other. Each step in our method resulted in greater overlap with the consensus set of genes predicted by both CANDID and MetaRanker (supplemental Table S2). However, given that the maximum overlap between any pairwise combination of the gene sets was 15% (CANDID and MetaRanker versus TC-GCM), we were still assured of the complementarity of each of the gene sets.

We further evaluated our results using DAPPLE (7) and GRAIL (38). DAPPLE looks for significant protein-protein interaction connectivity among proteins encoded by genes in loci associated with disease (7). GRAIL describes the degree of functional connectivity between regions using literature based relationships between genes (38). Our method had 31% similarity to DAPPLE prioritized candidate genes when the same seed genes were used, and 18% similarity to GRAIL results.

Because of GeneWanderer's top rating among network based approaches for gene prioritization (39), we used it to rank GCM genes with respect to the polygenic trait of obesity. GeneWanderer identified 48 of the 51 GCM genes as highly ranked candidates in genomic locations related to obesity. In our interactome, these 48 genes were immediate neighbors of genes within loci identified by GLGC GWAS ($p < 0.0001$) (supplemental Fig. S6).

Relevance of GCM Genes to Lipid Related Diseases Based on Literature—Many of the GCM candidate genes associated with all four traits have been linked to lipid metabolism, cardiovascular disease and coronary artery disease (supplemental Table S3). Retinoid x-receptor alpha (*RXRA*) variant

TABLE II

Prioritized co-GCM genes represented by SNPs in evolutionarily conserved regions for further genotyping in Malmö Diet and Cancer Cardiovascular cohort (MDC-CC) cohort

Trait	Gene	SNP	GLGC <i>p</i> -value	Variant type/location	DAPPLE	Genewanderer ranking	GRAIL <i>p</i> value
HDL-C	<i>INSR</i>	rs8101064	2.03E-05	INTRONIC	✓	1	0.9
	<i>ASCC2</i>	rs140147	0.0006	SPLICE_SITE		3	0.99
	<i>CYP3A4</i>	rs12721617	0.002	INTRONIC		1	0.031
	<i>APP</i>	rs380713	0.003	INTERGENIC		1	0.3
	<i>SMURF2</i>	rs6504248	0.006	INTERGENIC		1	0.96
	<i>EHMT2</i>	rs9267659	0.008	INTRONIC		7	4.92E-11
	<i>SKIL</i>	rs6763533	0.009	INTRONIC		1	0.96
	<i>PSMA1</i>	rs12362721	0.02	INTRONIC		3	0.126
	<i>TERT</i>	rs6554691	0.02	INTRONIC		1	0.636
	<i>RALYL</i>	rs6473532	0.03	INTERGENIC		NA	0.956
	<i>FASN</i>	rs6502051	0.04	INTRONIC		1	0.45
LDL-C	<i>NDUFA4L2</i>	rs11172134	0.0003	UPSTREAM	✓	NA	0.0006
	<i>CDK5RAP2</i>	rs3739822	0.0008	SYNONYMOUS_CODING	✓	5	0.65
	<i>ITGB3BP</i>	rs6588048	0.003	INTRONIC		3	0.98
	<i>SH3GL3</i>	rs8025427	0.03	INTRONIC		1	0.48
TC	<i>CBS</i>	rs234706	0.007	SYNONYMOUS_CODING		2	0.13
	<i>ASAP1</i>	rs7462286	0.02	INTRONIC	✓	1	0.42
	<i>ITSN1</i>	rs9984662	0.03	3PRIME_UTR		13	NA
	<i>EXOSC10</i>	rs11583740	0.03	INTRONIC		5	0.75
TG	<i>DNM2</i>	rs3826803	0.003	INTRONIC		5	0.85
	<i>HNF4A</i>	rs3212198	0.003	INTRONIC		1	0.22
	<i>PAFAH1B3</i>	rs3826706	0.02	INTRONIC	✓	5	0.7
	<i>COPS6</i>	rs2307345	0.02	INTRONIC		5	0.43
	<i>ATP6V1E1</i>	rs3532	0.02	3PRIME_UTR		3	0.86
	<i>NR2F2</i>	rs4310804	0.04	INTERGENIC		2	NA

rs11185660 has been associated with low HDL-C and coronary heart disease (40, 41). TG and nonesterified fatty acid (NEFA) levels were increased in the livers and serum of cystathionine-beta-synthase (*CBS*) knock out mice (41). Deletion of the four and a half LIM domains 2 (*FHL2*) gene attenuates the formation of atherosclerotic lesions normally present with a cholesterol-enriched diet (42).

Selection of SNPs for Genotyping in the MDC-CC—Because we expected elements of phenotype-specific modules to act cooperatively, we tested whether GCM genes were co-expressed with seed genes. Most of the GCM genes (90%) were significantly co-expressed with at least one of the seed genes (supplemental Table S1). Within the HDL-C, LDL-C, TC and TG GCM gene sets, 19, 9, 13 and 12 genes were co-expressed with seed genes having GWAS *p* values <0.05. As evolutionary conservation of genomic regions implies greater biological significance (30), we prioritized co-GCM genes represented by SNPs in evolutionarily conserved regions for further genotyping in MDC-CC cohort (Table III). The numbers of SNPs representing co-GCM genes in conserved regions for HDL-C, LDL-C, TC and TG traits were 11, 4, 4, and 6, respectively (Table II). The SNPs with the lowest GLGC GWAS *p* values in each of these gene groups were genotyped in the 5763 MDC-CC participants (Table III, Table IV).

TABLE III

Characteristics of the subjects in the Malmö Diet and Cancer Cardiovascular cohort

Clinical character	All (n = 5056)
Males/Females N (%)	2054 (40.6)/3002 (59.4)
Age (years)	57.5 ± 5.9
Body mass index (kg/m ²)	25.8 ± 3.9
HDL-cholesterol (mmol/l)	1.4 ± 0.4
LDL-cholesterol (mmol/l)	4.2 ± 1.0
Total cholesterol (mmol/l)	6.2 ± 1.1
Triglycerides (mmol/l)	1.4 ± 0.8
Diabetes N (%)	416 (8.2)

Logistic regression analysis of the four SNPs revealed that the minor A-allele of the synonymous SNP rs234706 in *CBS* (Y233Y) was significantly associated with higher total cholesterol levels than the G-allele (*p* = 0.013 after Bonferroni correction, Table IV). The A-allele also associated significantly with higher LDL-C (*p* = 0.00001) and TG (*p* = 0.04) levels. The three other SNPs did not associate significantly with their respective traits. Despite this, we found that the combined effect of all four risk-alleles was nominally significant for an association with lower HDL-cholesterol and higher triglyceride levels (*p* = 0.041 and 0.026, respectively, Supplemental Table 4). No evidence for pairwise epistasis between the SNPs was found.

TABLE IV

Association of selected GCM gene SNPs with their respective traits in MDC-CC. A synonymous SNP (rs234706) in CBS gene was significantly associated with the TC trait in MDC-CC

Trait	N	SNP (Chr)	Gene	Minor Allele (frequency, %)	Beta (S.E.)	p value
HDL-C	4916	rs8101064 (19)	INSR	T (3.9)	-0.001 (0.001)	0.50
LDL-C	4822	rs11172134 (12)	NDUFA4L2	A (23.7)	0.003 (0.02)	0.91
TC	4962	rs234706 (21)	CBS	A (45.9)	0.06 (0.02)	0.0032
TG	4933	rs3826803 (19)	DNM2	C (35.0)	0.004 (0.01)	0.68

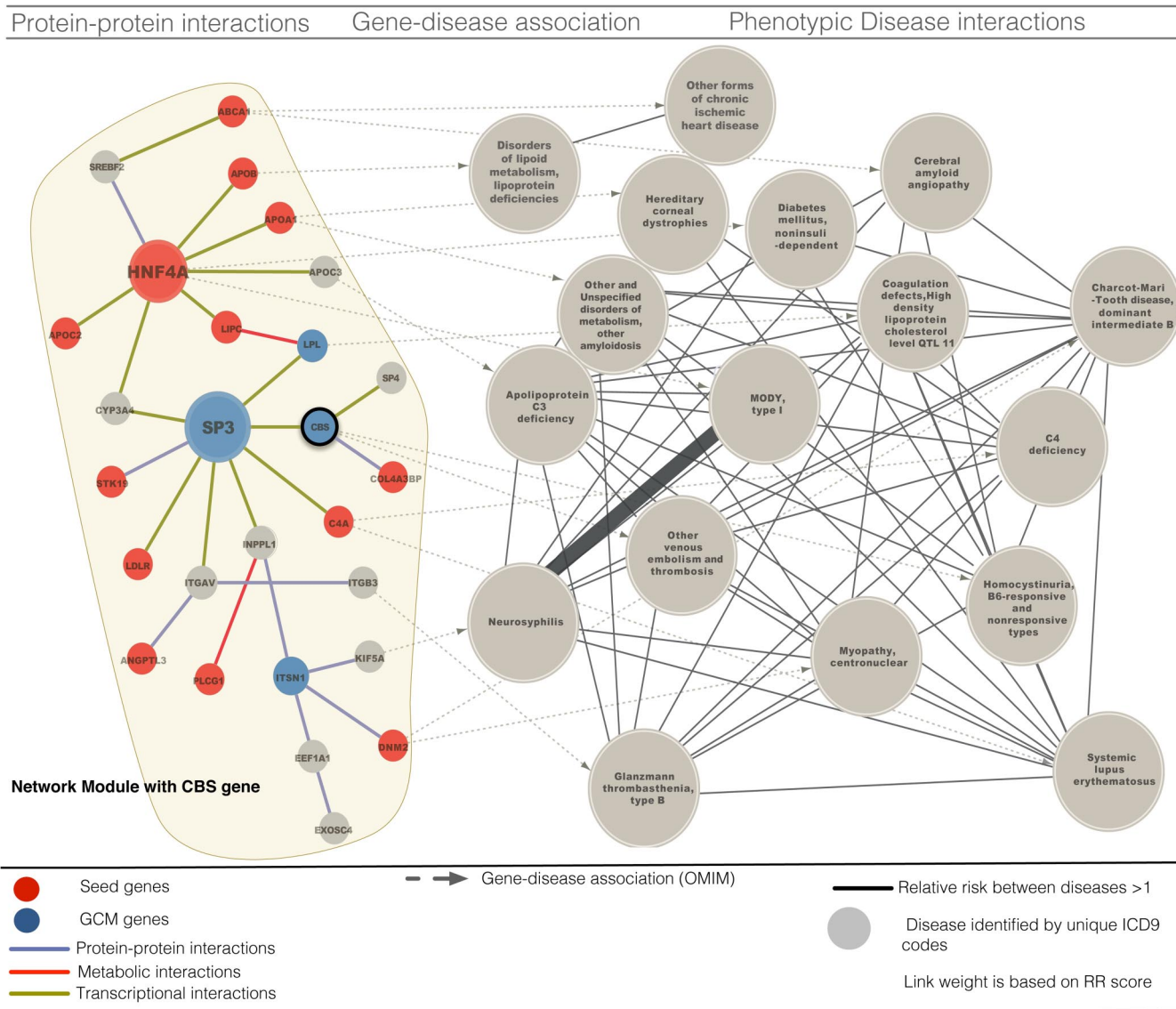


FIG. 5. **GCM module with CBS gene and the associated diseases.** Combination schema including protein-protein interactions (purple), metabolic interactions (red), and transcriptional interactions (yellow), gene-disease associations (dashed black), and relative risk associations between diseases greater with magnitude greater than 1 (black line). Seed genes (red ovals), CBS GCM genes (dark blue ovals) and diseases (gray) are linked within a highly interconnected module that includes Homocystinuria, venous embolism and thrombosis diseases associated with CBS gene in OMIM.

Expression of the CBS gene was associated with the directly measured rs234705 SNP, which served as a perfect proxy for rs234706 (HapMap CEU LD of $R^2 = 1$). The

rs234706 SNP was genotyped in the GLGC GWAS, and an association between CBS gene expression and the rs234706 genotype was determined (43). The minor allele of SNP

rs234706 was significantly associated with mRNA levels of CBS in the 206 liver biopsy samples ($p = 0.04$). In the disease to gene mappings, homocystinuria, venous embolisms, and thrombosis were associated with the CBS gene (Fig. 5). Coagulation defects, Diabetes mellitus and Charcot-Marie-Tooth disease were associated with other genes in the CBS GCM module, and comorbidities were found between these diseases (with all disease pairs having $RR > 1$, Fig. 5). We also found genes associated with homocystinuria and APOA1 associated amyloidosis within the CBS GCM module. These comorbid diseases have a RR score of 6.4, and the relationship between TC, CBS, homocystinuria, APOA1, and amyloidosis is supported by the observation that plasma cholesterol and APOA1 are significantly decreased in homozygous CBS-deficient mice (44).

DISCUSSION

Although the human interactome is far from complete, merging network topological features with heterogeneous GWAS data provides experimentally verifiable insights into complex biological traits. Unlike approaches that test genes in GWAS identified loci for overrepresentation in pathways (3), our approach uses network context to prioritize specific candidate genes. The improvement in the precision of our predictions, related to the high coverage of seed genes by our modules, is supported by coherent gene-disease and comorbidity associations. This highlights how seemingly unrelated diseases may be the product of complex combinations of shared molecular mechanisms. We believe that this allows our three step procedure to compete with more established methods such as CANDID and MetaRanker, and to capture additional candidate genes missed by other methods (Table II).

The GCM approach prospectively allows us to use nominally significant GWAS p values in the hunt for missing heritability while minimizing spurious hits. Many of our prioritized candidates for lipid traits are related to cardiovascular and coronary artery disease in the literature (40, 42, 45) (supplemental Table S3), providing a common sense measure of the usefulness of our raw data and methodology.

The significant association of the synonymous SNP rs234706 within the CBS gene to the TC and LDL-C traits, together with the association between the synonymous SNP and variable CBS mRNA levels in the liver (likely because of linkage disequilibrium with the gene's transcriptional regulatory elements (46)) suggests that CBS expression levels are related to aberrant lipid profile traits in humans. It has been shown that CBS knockout mice have altered distributions of cholesterol and triglyceride lipoprotein fractions, and that mutations in the CBS gene cause altered lipoprotein metabolism as well as hyperhomocysteinemia (47). This finding demonstrates the usefulness of the GCM approach in selecting lipid and lipoprotein trait associated candidate genes.

Population level disease comorbidity between genes revealed interconnected complex phenotypes. Integrating lipid

interactome data with patient medical records uncovered molecular associations for diseases unexpectedly comorbid with lipid related disorders. Despite the incompleteness of current protein-protein interactions and our incomplete knowledge of disease gene associations, the GCM method validated in one of the four SNPs tested. This 25% validation success rate surpasses that of other candidate gene prediction methods (8, 48, 49).

Although the GCM approach has been demonstrated using GWAS of lipid traits, it can be used to interpret GWAS of other traits as well. By capturing phenotypically coherent modules of candidate and seed genes, the GCM approach provides insights regarding involvement in complex phenotypes with multiple susceptibility alleles and low effect sizes. In this way, GCM as well as other network-based approaches may be of broad use in dissecting complex diseases in the coming era of systems medicine.

Acknowledgments—The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. We thank Sekar Kathiresan for providing access to Global Lipids Genetics Consortium, GLGC- GWAS data. We are thankful to lossifov for providing the program to run molecular train-gulation algorithm.

* This study is supported by the Swedish Research Council, the Swedish Heart and Lung Foundation, the Region Skåne, the Skåne University Hospital, the Novo Nordic Foundation, the Albert Pålsson Research Foundation, the Crafoord foundation equipment grant from the Knut and Alice Wallenberg Foundation and by Linnéus for the Lund University Diabetes Center (LUDC). MOM is a senior scientist at the Swedish Research Council. This work was supported by National Institutes of Health (NIH) grants P50-HG004233 CEGS.

§ This article contains supplemental Figs S1 to S6 and Tables S1 to S4.

§§ To whom correspondence should be addressed: Lund University, Skania University Hospital, CRC Entrance 72, Building 91 Floor 12, SE-205 02 Malmö, Sweden. Tel.: +46 40 39 12 10; Fax: +46 40 39 12 22; E-mail: amitabhsharma13@gmail.com, and marju.orholmeland@med.lu.se.

Conflict of interest: The authors declare that they have no conflict of interest.

REFERENCES

- Manolio, T. A., Collins, F. S., Cox, N. J., Goldstein, D. B., Hindorf, L. A., Hunter, D. J., McCarthy, M. I., Ramos, E. M., Cardon, L. R., Chakravarti, A., Cho, J. H., Guttmacher, A. E., Kong, A., Kruglyak, L., Mardis, E., Rotimi, C. N., Slatkin, M., Valle, D., Whittemore, A. S., Boehnke, M., Clark, A. G., Eichler, E. E., Gibson, G., Haines, J. L., Mackay, T. F., McCarroll, S. A., and Visscher, P. M. (2009) Finding the missing heritability of complex diseases. *Nature* **461**, 747–753
- Hegele, R. A. (2010) Genome-wide association studies of plasma lipids: have we reached the limit? *Arterioscler. Thromb. Vasc. Biol.* **30**, 2084–2086
- Holmans, P., Green, E. K., Pahwa, J. S., Ferreira, M. A., Purcell, S. M., Sklar, P., Owen, M. J., O'Donovan, M. C., and Craddock, N. (2009) Gene ontology analysis of GWA study data sets provides insights into the biology of bipolar disorder. *Am. J. Hum. Genet.* **85**, 13–24
- Wang, K., Zhang, H., Kugathasan, S., Annesse, V., Bradfield, J. P., Russell, R. K., Sleiman, P. M., Imielinski, M., Glessner, J., Hou, C., Wilson, D. C., Walters, T., Kim, C., Frackelton, E. C., Lionetti, P., Barabino, A., Van Limbergen, J., Guthery, S., Denson, L., Piccoli, D., Li, M., Dubinsky, M., Silverberg, M., Griffiths, A., Grant, S. F., Satsangi, J., Baldassano, R., and

- Hakonarson, H. (2009) Diverse genome-wide association studies associate the IL12/IL23 pathway with Crohn Disease. *Am. J. Hum. Genet.* **84**, 399–405
5. Zhong, H., Yang, X., Kaplan, L. M., Molony, C., and Schadt, E. E. (2010) Integrating pathway analysis and genetics of gene expression for genome-wide association studies. *Am. J. Hum. Genet.* **86**, 581–591
 6. Baranzini, S. E., Galwey, N. W., Wang, J., Khankhanian, P., Lindberg, R., Pelletier, D., Wu, W., Uitdehaag, B. M., Kappos, L., Polman, C. H., Matthews, P. M., Hauser, S. L., Gibson, R. A., Oksenberg, J. R., and Barnes, M. R. (2009) Pathway and network-based analysis of genome-wide association studies in multiple sclerosis. *Hum. Mol. Genet.* **18**, 2078–2090
 7. Rossin, E. J., Lage, K., Raychaudhuri, S., Xavier, R. J., Tatar, D., Benita, Y., Cotsapas, C., and Daly, M. J. (2011) Proteins encoded in genomic regions associated with immune-mediated disease physically interact and suggest underlying biology. *PLoS Genet.* **7**, e1001273
 8. Pers, T. H., Hansen, N. T., Lage, K., Koefoed, P., Dworzynski, P., Miller, M. L., Flint, T. J., Møllerup, E., Dam, H., Andreassen, O. A., Djurovic, S., Melle, I., Børglum, A. D., Werge, T., Purcell, S., Ferreira, M. A., Kouskoumvekaki, I., Workman, C. T., Hansen, T., Mors, O., and Brunak, S. (2011) Meta-analysis of heterogeneous data sources for genome-scale identification of risk genes in complex phenotypes. *Genet. Epidemiol.* **35**, 318–332
 9. Hutz, J. E., Kraja, A. T., McLeod, H. L., and Province, M. A. (2008) CANDID: a flexible method for prioritizing candidate genes for complex human traits. *Genet. Epidemiol.* **32**, 779–790
 10. Hartwell, L. H., Hopfield, J. J., Leibler, S., and Murray, A. W. (1999) From molecular to modular cell biology. *Nature* **402**, C47–52
 11. Goh, K. I., Cusick, M. E., Valle, D., Childs, B., Vidal, M., and Barabási, A. L. (2007) The human disease network. *Proc. Natl. Acad. Sci. U. S. A.* **104**, 8685–8690
 12. Krauthammer, M., Kaufmann, C. A., Gilliam, T. C., and Rzhetsky, A. (2004) Molecular triangulation: bridging linkage and molecular-network information for identifying candidate genes in Alzheimer's disease. *Proc. Natl. Acad. Sci. U. S. A.* **101**, 15148–15153
 13. Ideker, T., Ozier, O., Schwikowski, B., and Siegel, A. F. (2002) Discovering regulatory and signalling circuits in molecular interaction networks. *Bioinformatics* **18**, S233–240
 14. Hidalgo, C. A., Blumm, N., Barabási, A. L., and Christakis, N. A. (2009) A dynamic network approach for the study of human phenotypes. *PLoS Comput. Biol.* **5**, e1000353
 15. Lee, D. S., Park, J., Kay, K. A., Christakis, N. A., Oltvai, Z. N., and Barabási, A. L. (2008) The implications of human metabolic network topology for disease comorbidity. *Proc. Natl. Acad. Sci. U.S.A.* **105**, 9880–9885
 16. Teslovich, T. M., Musunuru, K., Smith, A. V., Edmondson, A. C., Stylianou, I. M., Koseki, M., Pirruccello, J. P., Ripatti, S., Chasman, D. I., Willer, C. J., Johansen, C. T., Fouchier, S. W., Isaacs, A., Peloso, G. M., Barbalic, M., Ricketts, S. L., Bis, J. C., Aulchenko, Y. S., Thorleifsson, G., Feitosa, M. F., Chambers, J., Orho-Melander, M., Melander, O., Johnson, T., Li, X., Guo, X., Li, M., Shin, Cho, Y., Jin, Go, M., Jin, Kim, Y., Lee, J. Y., Park, T., Kim, K., Sim, X., Twee-Hee, Ong, R., Croteau-Chonka, D. C., Lange, L. A., Smith, J. D., Song, K., Hua, Zhao, J., Yuan, X., Luan, J., Lamina, C., Ziegler, A., Zhang, W., Zee, R. Y., Wright, A. F., Witteman, J. C., Wilson, J. F., Willemssen, G., Wichmann, H. E., Whitfield, J. B., Waterworth, D. M., Wareham, N. J., Waeber, G., Vollenweider, P., Voight, B. F., Vitart, V., Uitterlinden, A. G., Uda, M., Tuomilehto, J., Thompson, J. R., Tanaka, T., Surakka, I., Stringham, H. M., Spector, T. D., Soranzo, N., Smit, J. H., Sinisalo, J., Silander, K., Sijbrands, E. J., Scuteri, A., Scott, J., Schlessinger, D., Sanna, S., Salomaa, V., Saharinen, J., Sabatti, C., Ruokonen, A., Rudan, I., Rose, L. M., Roberts, R., Rieder, M., Psaty, B. M., Pramstaller, P. P., Pichler, I., Perola, M., Penninx, B. W., Pedersen, N. L., Pattaro, C., Parker, A. N., Pare, G., Oostra, B. A., O'Donnell, C. J., Nieminen, M. S., Nickerson, D. A., Montgomery, G. W., Meitinger, T., McPherson, R., McCarthy, M. I., McArdle, W., Masson, D., Martin, N. G., Marroni, F., Mangino, M., Magnusson, P. K., Lucas, G., Luben, R., Loos, R. J., Lokki, M. L., Lettre, G., Langenberg, C., Launer, L. J., Lakatta, E. G., Laaksonen, R., Kyvik, K. O., Kronenberg, F., König, I. R., Khaw, K. T., Kaprio, J., Kaplan, L. M., Johansson, A., Jarvelin, M. R., Janssens, A. C., Ingelsson, E., Igl, W., Kees, Hovingh, G., Hottenga, J. J., Hofman, A., Hicks, A. A., Hengstenberg, C., Heid, I. M., Hayward, C., Havulinna, A. S., Hastie, N. D., Harris, T. B., Haritunians, T., Hall, A. S., Gyllenstein, U., Guiducci, C., Groop, L. C., Gonzalez, E., Gieger, C., Freimer, N. B., Ferrucci, L., Erdmann, J., Elliott, P., Ejebe, K. G., Döring, A., Dominiczak, A. F., Demissie, S., Deloukas, P., de Geus, E. J., de Faire, U., Crawford, G., Collins, F. S., Chen, Y. D., Caulfield, M. J., Campbell, H., Burt, N. P., Bonnycastle, L. L., Boomsma, D. I., Boekholdt, S. M., Bergman, R. N., Barroso, I., Bandinelli, S., Ballantyne, C. M., Assimes, T. L., Quertermous, T., Altshuler, D., Seielstad, M., Wong, T. Y., Tai, E. S., Feranil, A. B., Kuzawa, C. W., Adair, L. S., Taylor, H. A., Jr, Borecki, I. B., Gabriel, S. B., Wilson, J. G., Holm, H., Thorsteinsdottir, U., Gudnason, V., Krauss, R. M., Mohlke, K. L., Ordovas, J. M., Munroe, P. B., Kooner, J. S., Tall, A. R., Hegele, R. A., Kastelein, J. J., Schadt, E. E., Rotter, J. I., Boerwinkle, E., Strachan, D. P., Mooser, V., Stefansson, K., Reilly, M. P., Samani, N. J., Schunkert, H., Cupples, L. A., Sandhu, M. S., Ridker, P. M., Rader, D. J., van, Duijn, C. M., Peltonen, L., Abecasis, G. R., Boehnke, M., Kathiresan, S.). (2010) Biological, clinical and population relevance of 95 loci for blood lipids. *Nature* **466**, 707–713
 17. Hong, M. G., Pawitan, Y., Magnusson, P. K., and Prince, J. A. (2009) Strategies and issues in the detection of pathway enrichment in genome-wide association studies. *Hum. Genet.* **126**, 289–301
 18. Rual, J. F., Venkatesan, K., Hao, T., Hirozane-Kishikawa, T., Dricot, A., Li, N., Berriz, G. F., Gibbons, F. D., Dreze, M., Ayivi-Guedehoussou, N., Klitgord, N., Simon, C., Boxem, M., Milstein, S., Rosenberg, J., Goldberg, D. S., Zhang, L. V., Wong, S. L., Franklin, G., Li, S., Albal, J. S., Lim, J., Fraughton, C., Llamosas, E., Cevik, S., Bex, C., Lamesch, P., Sikorski, R. S., Vandenhaute, J., Zoghbi, H. Y., Smolyar, A., Bosak, S., Sequerra, R., Doucette-Stamm, L., Cusick, M. E., Hill, D. E., Roth, F. P., and Vidal, M. (2005) Towards a proteome-scale map of the human protein-protein interaction network. *Nature* **437**, 1173–1178
 19. Stelzl, U., Worm, U., Lalowski, M., Haenig, C., Brembeck, F. H., Goehler, H., Stroedicke, M., Zenkner, M., Schoenherr, A., Koepfen, S., Timm, J., Mintzlaff, S., Abraham, C., Bock, N., Kietzmann, S., Goedde, A., Toksoz, E., Droege, A., Krobitsch, S., Korn, B., Birchmeier, W., Lehrach, H., and Wanker, E. E. (2005) A human protein-protein interaction network: a resource for annotating the proteome. *Cell* **122**, 957–968
 20. Venkatesan, K., Rual, J. F., Vazquez, A., Stelzl, U., Lemmens, I., Hirozane-Kishikawa, T., Hao, T., Zenkner, M., Xin, X., Goh, K. I., Yildirim, M. A., Simonis, N., Heinzmann, K., Gebreab, F., Sahalie, J. M., Cevik, S., Simon, C., de Smet, A. S., Dann, E., Smolyar, A., Vinayagam, A., Yu, H., Szeto, D., Borick, H., Dricot, A., Klitgord, N., Murray, R. R., Lin, C., Lalowski, M., Timm, J., Rau, K., Boone, C., Braun, P., Cusick, M. E., Roth, F. P., Hill, D. E., Tavernier, J., Wanker, E. E., Barabási, A. L., and Vidal, M. (2009) An empirical framework for binary interactome mapping. *Nat. Methods* **6**, 83–90
 21. Ceol, A., Chatr-Aryamontri, A., Licata, L., Peluso, D., Briganti, L., Perfetto, L., Castagnoli, L., and Cesareni, G. (2010) MINT, the molecular interaction database: 2009 update. *Nucleic Acids Res.* **38**, D532–539
 22. Aranda, B., Achuthan, P., Alam-Farouque, Y., Armean, I., Bridge, A., Derow, C., Feuermann, M., Ghanbarian, A. T., Kerrien, S., Khadake, J., Kerssemakers, J., Leroy, C., Menden, M., Michaut, M., Montecchi-Palazzi, L., Neuhauser, S. N., Orchard, S., Perreau, V., Roehert, B., van Eijk, K., and Hermjakob, H. (2010) The IntAct molecular interaction database in 2010. *Nucleic Acids Res.* **38**, D525–531
 23. Matys, V., Kel-Margoulis, O. V., Fricke, E., Liebich, I., Land, S., Barre-Dirrie, A., Reuter, I., Chekmenev, D., Krull, M., Hornischer, K., Voss, N., Stegmaier, P., Lewicki-Potapov, B., Saxel, H., Kel, A. E., and Wingender, E. (2006) TRANSFAC and its module TRANSCOMP: transcriptional gene regulation in eukaryotes. *Nucleic Acids Res.* **34**, D108–110
 24. Iossifov, I., Rodriguez-Esteban, R., Mayzus, I., Millen, K. J., and Rzhetsky, A. (2009) Looking at cerebellar malformations through text-mined interactomes of mice and humans. *PLoS Comput. Biol.* **5**, e1000559
 25. Zheng, S., and Zhao, Z. (2012) GenRev: Exploring functional relevance of genes in molecular networks. *Genomics* **99**, 183–188
 26. Sun, P. G., Gao, L., and Han, S. (2011) Prediction of human disease-related gene clusters by clustering analysis. *Int. J. Biol. Sci.* **7**, 61–73
 27. Schlicker, A., Lengauer, T., and Albrecht, M. (2010) Improving disease gene prioritization using the semantic similarity of Gene Ontology terms. *Bioinformatics* **26**, i561–567
 28. Köhler, S., Bauer, S., Horn, D., and Robinson, P. N. (2008) Walking the interactome for prioritization of candidate disease genes. *Am. J. Hum. Genet.* **82**, 949–958
 29. Su, A. I., Wiltshire, T., Batalov, S., Lapp, H., Ching, K. A., Block, D., Zhang,

- J., Soden, R., Hayakawa, M., Kreiman, G., Cooke, M. P., Walker, J. R., and Hogenesch, J. B. (2004) A gene atlas of the mouse and human protein-encoding transcriptomes. *Proc. Natl. Acad. Sci. U.S.A.* **101**, 6062–6067
30. Adzhubei, I. A., Schmidt, S., Peshkin, L., Ramensky, V. E., Gerasimova, A., Bork, P., Kondrashov, A. S., and Sunyaev, S. R. (2010) A method and server for predicting damaging missense mutations. *Nat. Methods* **7**, 248–249
31. Reumers, J., Conde, L., Medina, I., Maurer-Stroh, S., Van Durme, J., Dopazo, J., Rousseau, F., and Schymkowitz, J. (2008) Joint annotation of coding and non-coding single nucleotide polymorphisms and mutations in the SNPeffect and PupaSuite databases. *Nucleic Acids Res.* **36**, D825–829
32. Berglund, G., Elmstahl, S., Janzon, L., and Larsson, S. A. (1993) The Malmo Diet and Cancer Study. Design and feasibility. *J. Intern Med.* **233**, 45–51
33. Jerntorp, P., and Berglund, G. (1992) Stroke registry in Malmo, Sweden. *Stroke* **23**, 357–361
34. Folkersen, L., Wagsater, D., Paloschi, V., Jackson, V., Petrini, J., Kurtovic, S., Maleki, S., Eriksson, M. J., Caidahl, K., Hamsten, A., Michel, J. B., Liska, J., Gabrielsen, A., Franco-Cereceda, A., and Eriksson, P. (2011) Unraveling the divergent gene expression profiles in bicuspid and tricuspid aortic valve patients with thoracic aortic dilatation - the ASAP study. *Mol. Med.* **17**, 1365–1373
35. Maere, S., Heymans, K., and Kuiper, M. (2005) BiNGO: a Cytoscape plugin to assess overrepresentation of gene ontology categories in biological networks. *Bioinformatics* **21**, 3448–3449
36. Barabási, A. L., Gulbahce, N., and Loscalzo, J. (2011) Network medicine: a network-based approach to human disease. *Nat. Rev. Genet.* **12**, 56–68
37. Scott, M. S., Perkins, T., Bunnell, S., Pepin, F., Thomas, D. Y., and Hallett, M. (2005) Identifying regulatory subnetworks for a set of genes. *Mol. Cell. Proteomics* **4**, 683–692
38. Raychaudhuri, S., Plenge, R. M., Rossin, E. J., Ng, A. C., Purcell, S. M., Sklar, P., Scolnick, E. M., Xavier, R. J., Altshuler, D., and Daly, M. J. (2009) Identifying relationships among genomic disease regions: predicting genes at pathogenic SNP associations and rare deletions. *PLoS Genet.* **5**, e1000534
39. Navlakha, S., and Kingsford, C. (2010) The power of protein interaction networks for associating genes with diseases. *Bioinformatics* **26**, 1057–1063
40. Peloso, G. M., Demissie, S., Collins, D., Mirel, D. B., Gabriel, S. B., Cupples, L. A., Robins, S. J., Schaefer, E. J., and Brousseau, M. E. (2010) Common genetic variation in multiple metabolic pathways influences susceptibility to low HDL-cholesterol and coronary heart disease. *J. Lipid Res.* **51**, 3524–3532
41. Namekata, K., Enokido, Y., Ishii, I., Nagai, Y., Harada, T., and Kimura, H. (2004) Abnormal lipid metabolism in cystathionine beta-synthase-deficient mice, an animal model for hyperhomocysteinemia. *J. Biol. Chem.* **279**, 52961–52969
42. Chu, P. H., Yeh, H. I., Wu, H. H., Hong, R. C., Shiu, T. F., and Yang, C. M. (2010) Deletion of the FHL2 gene attenuates the formation of atherosclerotic lesions after a cholesterol-enriched diet. *Life Sci.* **86**, 365–371
43. Folkersen, L., van't Hooft, F., Chernogubova, E., Agardh, H. E., Hansson, G. K., Hedin, U., Liska, J., Syvanen, A. C., Paulsson-Berne, G., Franco-Cereceda, A., Hamsten, A., Gabrielsen, A., and Eriksson, P. (2010) Association of genetic risk variants with expression of proximal genes identifies novel susceptibility genes for cardiovascular disease. *Circ. Cardiovasc Genet.* **3**, 365–373
44. Nuño-Ayala, M., Guillen, N., Navarro, M. A., Lou-Bonafonte, J. M., Arnal, C., Gascon, S., Barranquero, C., Godino, J., Royo-Canas, M., Sarria, A. J., Guzman, M. A., Hernandez, E., Bregante, M. A., Garcia-Gimeno, M. A., and Osada, J. (2010) Cysteinemia, rather than homocysteinemia, is associated with plasma apolipoprotein A-I levels in hyperhomocysteinemia: lipid metabolism in cystathionine beta-synthase deficiency. *Atherosclerosis* **212**, 268–273
45. Palanker, L., Tennessen, J. M., Lam, G., and Thummel, C. S. (2009) Drosophila HNF4 regulates lipid mobilization and beta-oxidation. *Cell Metab.* **9**, 228–239
46. Aras, O., Hanson, N. Q., Yang, F., and Tsai, M. Y. (2000) Influence of 699C->T and 1080C->T polymorphisms of the cystathionine beta-synthase gene on plasma homocysteine levels. *Clin. Genet.* **58**, 455–459
47. Liao, D., Tan, H., Hui, R., Li, Z., Jiang, X., Gaubatz, J., Yang, F., Durante, W., Chan, L., Schafer, A. I., Pownall, H. J., Yang, X., and Wang, H. (2006) Hyperhomocysteinemia decreases circulating high-density lipoprotein by inhibiting apolipoprotein A-I Protein synthesis and enhancing HDL cholesterol clearance. *Circ. Res.* **99**, 598–606
48. Tremblay, K., Lemire, M., Potvin, C., Tremblay, A., Hunninghake, G. M., Raby, B. A., Hudson, T. J., Perez-Iratxeta, C., Andrade-Navarro, M. A., and Laprise, C. (2008) Genes to diseases (G2D) computational method to identify asthma candidate genes. *PLoS One* **3**, e2907
49. Erlich, Y., Edvardson, S., Hodges, E., Zenvirt, S., Thekkat, P., Shaag, A., Dor, T., Hannon, G. J., and Elpeleg, O. (2011) Exome sequencing and disease-network analysis of a single family implicate a mutation in KIF1A in hereditary spastic paraparesis. *Genome Res.* **21**, 658–664